Effects of Chemically Pretreated Bovine Serum Samples on Sensitivity of Rose Bengal Test for Serodiagnosis of Bovine Brucellosis

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ABSTRACT

The standard serological test such as Rose Bengal Plate Test (RBPT) is routinely used for the diagnosis of brucellosis. This test depends on the agglutination of colored particulate antigen (killed *Brucella* organisms) by the antibodies present in sera of infected animals. Faulty negative and positive results are commonly experienced in these traditional agglutination tests. We developed three simple, new, additional and cost-effective steps that can help in these problems; Superagglutination test for serodiagnosis of brucellosis differs from conventional RBPT by three simple new and coast effective additional steps which are used to overcome this problem. These steps depend on the staining of sera antibodies by adding dye before the test and addition of diluted biotinylated antiglobulin and Avidin in sequence then mixing the antigen with the stained serum. By testing 150 serum samples, Superagglutination test had higher positive predictive value and specificity than RBPT and standard tube agglutination test (STAT). Also, it had higher negative predictive value and sensitivity than RBPT, STAT, ELISA (Indirect enzyme-linked immunosorbent assay) and CFT (Complement Fixation test).

Keywords: *Brucella*, CFT, ELISA, RBPT, STAT.

Original Article:

DOI: <https://dx.doi.org/10.21608/javs.2020.85550>

Received : 20 Jan., 2020

Accepted: 25 Feb., 2020

Published in April 2020

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J. Appl. Vet. Sci., 5(2) : 6-12.

INTRODUCTION

Brucellosis is an important worldwide zoonotic bacterial disease caused by *Brucella* species and affecting different mammals including man, sheep, goats, cattle, swine, marine mammals and rodents. The disease mainly affects the reproductive system with significant loss of productivity and re-productivity of affected animals. Control of brucellosis depends mainly on comprehensive vaccination, surveillance and quarantine programs. Both control and prevention procedures are highly depending on accurate diagnostic tools and effective and safe vaccination programs (WHO, 2006).

The RBPT is a pilot, cheap, rapid and effective screening test for *Brucella* diagnosis. It can be performed with minimum facilities. Because of its apparent simplicity, high level of standardization of antigen but the accuracy of the reading is needed

(Malek *et al.*, 2013). However, it may give false negative and positive results. Many factors affect its reaction and reading as some people can see the finer agglutination while many others cannot. Very low concentration of antibodies may not give visible agglutination. False-positive results may arise due to the inability to differentiate non-specific aggregates of antigen particles alone from the true agglutinates comprising both antigen and antibody. False-negative results may be due to a small clump size in sera with low titers of antibody. Prozone may also lead to the false-negative reaction when sera of high antibody titers are tested.

So, to get an accurate diagnosis of *Brucella* infection, a combination of RBPT, ELISA and CFT should be used. This trial aimed to develop a new Super agglutination test with higher positive and negative predictive values and elevated specificity and sensitivity than traditional RBPT to give more accurate

serodiagnosis without any change in simplicity and ease of test application and without increasing the cost of the test.

MATERIALS AND METHODS

1. Brucella strains

Strain-99 (S-99), smooth *B.abortus* biovar 1 (Weybridge, England) .

2. Brucella antigens for serologic tests

Conventional Rose Bengal and Tube Agglutination Antigens prepared from *B.abortus* (S-99) according to (Alton *et al.*, 1988 and OIE, 2016).

3. Serum samples

A total of one hundred-fifty cow's serum samples were collected from different farms in different Egyptian governorates.

4. Chemicals and Stains

4.1. Rose Bengal stain (Sigma) used for staining of Brucella cells (*B.abortus* S.99) according to Alton *et al.* (1988).

4.2. Coomassie Blue Stain (Sigma) used for staining of protein used for staining of serum in Superagglutination Test.

4.3. Biotinylated anti-bovine IgG (Sigma).

4.4. Streptavidin. (Sigma).

5. Serologic tests

5.1. Conventional Rose Bengal Plate Test (RBPT)

It was performed as described by Alton *et al.* (1988), briefly, the sera and antigen were brought to room temperature before use. Equal volumes (30µl) of standardized *B. abortus* antigen Weybridge strain 99 and test serum were mixed thoroughly and rotated on a clean glass slide using a stick applicator, and the slide was rocked for 4 min. The formation of clear clumps was considered a positive test while the absence of clear clumps was considered a negative reaction.

5.2. Standard Tube Agglutination test (STAT)

It was performed, following the procedure described by Alton *et al.* (1988), a plain antigen of Brucella abortus(S-99) was used. Two-fold serial dilutions (1:20 to 1:640) of the sera were prepared in phenol saline and equal volume (0.5ml) of antigen was added to each tube. All the tubes were incubated at 37°C for 24h. The results were compared with the antigen control tube showing 50%agglutination. A titer of 1:40 or above was considered positive.

5.3. Superagglutination Test (Modified Rose Bengal Plate Antigen (RBPA)

An equal volume (2.5 µl) of RBPA colored antigens and stained tested sera were mixed with

1µl of 0.1%Coomassie Blue dye, 1µl of biotinylated anti-bovine IgG and 1µl streptavidin, on a clean glass slide. The slide was observed for 4 minutes for the formation of clumps. Ordinary hand lens was used occasionally for better visibility. The slides were viewed under a low power (4×or 10X) objective lens under a microscope to confirm the clumping in case of doubt. Formation of clear clumps, within which presence of both blue colour and the pink colour together were considered as positive, while the absence of clear agglutinates or aggregates of pink color alone or blue-colored mass alone were considered as negative.

5.4. Indirect enzyme-linked immunosorbent assay (ELISA)

It was performed according to Alton *et al.*, (1988).

6. Complement Fixation test (CFT)

Which was performed on a microplate, following the procedure described by Alton *et al.* (1988). Complement fixation at a dilution of 3log2 (1:8), the level recommended by the Australian Bureau of Animal Health, (1979) which was regarded as a positive reaction. Serum samples were titrated 1:4 to 1:128 in the CFT. Titers determined by CFT were expressed as log2 of the reciprocal of the last dilution at which a positive reaction occurred (Australian Bureau of Animal Health, 1979).

Considering CFT as a gold standard test (Yohannes *et al.*, 2012) antigens sensitivity, relative sensitivity and specificity were calculated using (<http://vassarstats.net/clin1.html>) as shown in table (1).

Table 1: Calculation of sensitivity and specificity concerning gold standard test

		Gold Standard Test (Cft)		Total
		Positive	Negative	
Test Under Evaluation	Positive	A	B	A+B
	Negative	C	D	C+D
Total		A+C	B+D	N (264)

RelativeSensitivity=A/A+C

Specificity= D/D+B

True positive (Positive Predictive Value)=A/A+B

False-positive (B) =B/A+B

True negative (Negative Predictive Value) =D/D+C

False-negative (C) =C/D+C

RESULTS

In the present study, 150 bovine sera samples were tested by RBPT, STAT, ELISA, Modified Rose Bengal Antigen Superagglutination test and CFT as shown in table(2). Results revealed that there are no satisfactory differences observed in the specificity of all used tests (Table3). Concerning CFT as a gold standard test, satisfactory differences were observed insensitivity of different tests used in this study. The sensitivity of RBT, STAT and ELISA were 24.66 %, 26.66% and 28%, respectively, where the modified Rose Bengal test showed the highest sensitivity (31.33

%). Results of relative sensitivity showed a satisfactory difference between different tests with highest relative sensitivity in case of using Super-agglutination antigen (95.92%) where the relative sensitivity of other tests were 75.51 %, 81.63% and 85.71% using RBT, STAT and ELISA respectively (Table 3). Elevation in sensitivity is related to decrease in the number of false-negative cases which were 4.08%, 24.49%, 18.37% and 14.29% in Modified Rose Bengal test (Superagglutination), RBT, STAT and ELISA respectively as shown in tables (3, 4).

Table 2: Numbers of positive and negative samples in each conducting test

Serological test	RBPT		STAT		ELISA		Superagglutination test		CFT Gold Standard (CFT Test)	
Result	(+ve))	(-ve)	(+ve)	(-ve)	(+ve)	(-ve)	(+ve)	(-ve)	(+ve)	(-ve)
	37	113	40	110	42	108	47	103	49	101
Total	150		150		150		150		150	

Table (3): Relative Sensitivity and specificity of RBT, STAT, ELISA and Superagglutination test for diagnosis of bovine brucellosis

Tests	Sensitivity	Relative sensitivity	Specificity	True positive predictive	False-positive	True negative predictive	False-negative
RBPT	24.66%	75.51%	100%	100%	Zero	89.38%	24.49%
STAT	26.66%	81.63%	100%	100%	Zero	91.82%	18.37%
ELISA	28%	85.71%	100%	100%	Zero	93.52%	98.16%
Superagglutinationn test	31.33%	95.92%	100%	100%	Zero	98.06%	4.08%

Table 4: Results of RBT, STAT, ELISA and Superagglutination test against CFT

Test		CFT Gold Standard Test	
RBPT	Positive	37	0
	Negative	12	101
STAT	Positive	40	0
	Negative	9	101
ELISA	Positive	42	0
	Negative	7	101
Superagglutination test	Positive	47	0
	Negative	2	101

49 sample were positive
101 samples were negative

Fig.1: Macroscopic view of the slide in positive RBPT

Fig.2: Macroscopic view of the slide in positive and negative RBPT.

Fig.3: Microscopic view of the slide (low power) in negative RBPT.

Fig.4: Microscopic view of the slide (low power) in positive RBPT.

Fig.5: Macroscopic view of the slide in positive Superagglutination test.

Fig.6: Microscopic view of the slide (low power) in positive Superagglutination test.

Fig.7: Microscopic view of the slide (low power) in positive Superagglutination test.

Fig.8: Macroscopic view of the slide in the negative Superagglutination test.

Fig.9: Microscopic view of the slide (low power) in negative Superagglutination test.

DISCUSSION

Brucellosis is an important and widely prevalent zoonotic disease of human and animals caused by *Brucella* species (OIE, 2016). The most common, screening and rapid serological test used for the diagnosis of brucellosis is Rose Bengal plate test (RBPT) (Novita, 2017) which based on the agglutination of antigen (killed *Brucella abortus* S99) by the antibodies present in animals sera. Although it is a cheap, simple and effective test and it can be carried out with the minimum of equipment, and the result is read by the naked eye, the RBPT is generally considered to be less sensitive than other serological tests like standard tube agglutination test, complement fixation test (CFT) and ELISA (Chachra et al., 2009). Generally, the negative results given by RBPT and STAT when used alone need to be further confirmed by other tests like CFT and ELISA, to avoid any possibility of false diagnosis of brucellosis (Raies et al., 2013).

This study aimed to minimize false-negative results of RBPT during the screening of bovine brucellosis and overcome the prozone phenomena. Three simple steps were developed to overcome this problem. The first step was the addition of Coomassie blue which is a protein stain of contrasting color, when serum sample mixed with its serum antibodies stained blue. Then adding of biotinylated anti-bovine IgG and streptavidin to enhance the clump size by cross-linking the antibodies molecules making larger and more compact mass of clumps; it makes the clumps 4 times larger and compact. So, the Superagglutination test detected more positive samples than other serological tests, it identified all the sera giving false-negative results as well as sera giving false-positive results by

RBPT, STAT and ELISA. It could detect all the positive samples that were undetected by the other tests. The new Superagglutination test and common serological tests i.e. RBPT, STAT, ELISA and CFT were applied on all 150 serum samples. (Table 2). Formation of clear agglutination of both blue and pink color together could be considered as positive in case of Superagglutination test (figure 5,6,7), while the absence of clear agglutination and aggregation of pink color alone or blue-colored mass alone were considered as negative (figure 8,9).

The Superagglutination test gave superior results in detecting anti-brucella antibodies compared to the conventional rose bengal plate test (Abou-Chaker et al., 2009) which showed non-specific one colored aggregates by the naked eye (figure 1,2) and under low power of light microscope as in figure (3,4). Complement fixation tests were considered as a gold standard test to determine the sensitivity and specificity of tested antigens in absence of bacteriological isolation (Yohannes et al., 2012). Statistics in this study were considered the 95 % confidence intervals (95% CI).

Concerning CFT as a gold standard test, results of this study was in agreement with Tsai et al., (1991); Magnarelli et al., (1991); Chang, et al., (1993); Hu et al., (2014); Saxena et al., (2013 and 2015) in terms of that Superagglutination test showed the highest sensitivity when compared with other tests performed in this study which was (95.92%). This may be due to the lesser number of false-negative results obtained with Super agglutination test (4.08%) compared to the other tests as 2 samples only were negative by modified Rose Bengal antigen out of 49 sample (table 3). Whereas, there were 12, 9 and 7 negative samples

of brucellosis sera with traditional Rose Bengal test, STAT and ELISA tests respectively (table 4).

CONCLUSION

In Superagglutination test detected more positive samples than other serological tests, it could identify all the sera giving false-negative results as well as sera giving false-positive results by RBPT, STAT and ELISA. It could detect all the positive samples that were undetected by the other tests. Superagglutination test is simple and significantly enhance the sensitivity of the agglutination test. The Results of this study recommended the use of Superagglutination test as a good diagnostic and confirmatory test to be used for diagnosis of bovine brucellosis but more studies are needed to be done on large cattle populations accompanied with bacteriological isolation to detect the sensitivity and specificity of this test and also to validate it.

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How to cite this article:

Lamees A. El-Tantawy, Lubna, F. Farahat, Eman S. Ramadan, and Fatma F. Warda. 2020. Effects Of Chemically Pretreated Bovine Serum Samples On Sensitivity Of Rose Bengal Test For Serodiagnosis Of Bovine Brucellosis. *Journal of Applied Veterinary Sciences*, 5(2): 6-12.